

# Huperzine A: A promising anticonvulsant, disease modifying, and memory enhancing treatment option in Alzheimer's disease



Ugur Damar<sup>a</sup>, Roman Gersner<sup>a</sup>, Joshua T. Johnstone<sup>b</sup>, Steven Schachter<sup>c</sup>, Alexander Rotenberg<sup>a,\*</sup>

<sup>a</sup> F.M. Kirby Neurobiology Center, Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA

<sup>b</sup> Biscayne Pharmaceuticals, Inc., Miami, FL, USA

<sup>c</sup> Department of Neurology, Beth Israel Deaconess Medical Center, and Massachusetts General Hospital, Harvard Medical School, Boston, MA, USA

## ARTICLE INFO

### Article history:

Received 5 October 2016

Accepted 17 December 2016

### Keywords:

Huperzine A  
Acetylcholinesterase  
Nicotinic  
Seizure  
Alzheimer's disease  
Antiepileptic  
ppTMS

## ABSTRACT

Alzheimer's disease (AD) is the most frequent cause of dementia. Besides cognitive deterioration, patients with AD are prone to seizures – more than 20% of patients diagnosed with AD experience at least one unprovoked seizure and up to 7% have recurrent seizures. Although available antiepileptic drugs (AEDs) may suppress seizures in patients with AD, they may also worsen cognitive dysfunction and increase the risk of falls. On the basis of preclinical studies, we hypothesize that Huperzine A (HupA), a safe and potent acetylcholinesterase (AChE) inhibitor with potentially disease-modifying qualities in AD, may have a realistic role as an anticonvulsant in AD.

© 2016 Elsevier Ltd. All rights reserved.

## Introduction

Alzheimer's disease (AD) is the most frequent cause of dementia worldwide, and remains incurable. FDA-approved drugs to relieve AD symptoms are acetylcholinesterase (AChE) inhibitors and memantine, a noncompetitive N-methyl-D-aspartate (NMDA) antagonist. AChE inhibitors address mnemonic symptoms referable to basal forebrain and cortical cholinergic deafferentation in AD [1]. Memantine limits neuronal glutamate excitotoxicity, interferes with pathological NMDA-mediated glutamate signaling, and increases normal signal-to-noise ratio [2–4].

### *Epilepsy, its treatment in AD and associated drawbacks*

10–22% of AD patients have at least one unprovoked seizure [5] and 4–7% experience recurrent seizures [6–8]. Genetic factors and disease severity correlate significantly with AD epileptogenesis. Particularly patients with early-onset familial AD and patients with more severe AD symptoms are most likely to develop seizures [9]. Studies suggest that epileptiform activity in cortical and hippocampal network in mouse AD models decreases synaptic plasticity in dentate gyrus, which has been linked to cognitive decline in

AD [10,11]. Hence, one could hypothesize that prevention of seizures in AD could stabilize and perhaps even reverse cognitive impairment.

Numerous antiepileptic drugs (AEDs) have been tested in AD animal models and AD patients, but none have ideal clinical profiles. Preclinical data are largely derived from studies in transgenic rodent models. For instance, levetiracetam reduces abnormal spike-wave activity, and in chronic use (12 days), reverses hippocampal remodeling and cognitive deficits in mice [12], benzodiazepines (BZ) increase inhibitory transmission and ameliorate enhanced synaptic potentiation in AD [13], and valproate may decrease beta amyloid (A $\beta$ ) production, and thereby provide neuroprotection and facilitate cognitive improvement [14,15].

In contrast to the encouraging animal work, human trials have not produced strong support for chronic use of conventional AEDs in AD. One study showed that levetiracetam, lamotrigine, and phenobarbital provided seizure freedom in only 30% of patients with AD, and reduced seizure frequency by >50% in 42, 35, and 36% of the patients, respectively [16]. Yet levetiracetam, lamotrigine, and phenobarbital caused frequent adverse effects, i.e. headache, dizziness, asthenia and somnolence, in 17, 28 and 43% of patients, respectively. While levetiracetam treatment corresponded to cognitive improvement in AD patients, in addition to seizure control, it caused somnolence and increased risk for infection [16,17]. Other AEDs such as phenytoin, topiramate, valproate and lacosamide have a considerable range of adverse effects with chronic

\* Corresponding author at: 300 Longwood Ave, Department of Neurology, Boston Children's Hospital, Boston, MA 02115, USA.

E-mail address: [Alexander.Rotenberg@childrens.harvard.edu](mailto:Alexander.Rotenberg@childrens.harvard.edu) (A. Rotenberg).

use that renders them as suboptimal treatment options [16,18,19]. BZ use is also often impractical in the elderly due to the risk of sedation and falls [20] as well as BZ-related cognitive decline [21,22]. Overall, AED adverse effects are reported by about 34% of patients, and this figure almost doubles when written questionnaires are used to survey for symptoms [23]. Such adverse events lead to a withdrawal of AED treatment in about 25% of AD patients [24], which underscores a need for a novel therapeutic with a favorable side-effect profile in this syndrome.

#### *Epileptogenic pathology in AD*

Hippocampal sclerosis (HS) and A $\beta$  deposition, the two major culprits in AD pathology, increase seizure propensity [25]. HS is accompanied by regional reduction of neuropeptide Y (NPY) which has anti-epileptic properties [26]. Also, NPY-neurogliaform interneurons which are GABAergic and have dense cholinergic input are reduced in number in rat brains after pilocarpine-induced status epilepticus which causes HS-like damage [27,28]. A $\beta$  and neurofibrillary tangles (NFTs, excessively phosphorylated tau protein or paired helical filaments) are hallmarks of AD pathogenesis [29]. A $\beta$  spontaneously assembles into dimers, oligomers, and fibrils. Intracellular A $\beta$  (mainly A $\beta$ 42) and extracellular/secreted A $\beta$  (mainly A $\beta$ 40) contribute significantly to A $\beta$  toxicity [30]. A $\beta$ 42 forms amyloid fibrils faster than A $\beta$ 40, comprises the majority of amyloid plaques, and relates more intimately to AD pathology [31]. In vitro, cortical neurons from mice with mutant human amyloid precursor protein (hAPP) were exposed to extracellular A $\beta$ , particularly fibrillar A $\beta$ 1–42 or A $\beta$ 25–35 show increased pyramidal cell membrane excitability and predispose mice to seizures [32]. Studies conducted in transgenic mice overexpressing hAPP also show that an increase in A $\beta$  is sufficient to trigger spontaneous non-convulsive seizures recorded from cortex and hippocampus, even without overt neurodegeneration, suggesting that high A $\beta$  levels directly cause disrupted network synchronization [11,25,33].

A $\beta$  toxicity is dependent on the tau protein, which is phosphorylated and modulated by the Fyn tyrosine kinase [34]. While tau protein is necessary for tubulin to assemble into microtubules, hyperphosphorylated tau, as in AD, binds normal tau and microtubule-associated proteins and sequesters them into insoluble filament tangles, thereby inhibiting microtubule assembly [35]. Tau hyperphosphorylation has been associated with excitotoxicity, neurodegeneration, and various causes of epilepsy such as traumatic brain injury and focal cortical dysplasia [36,37]. In further support of the tau role in AD epileptogenesis, genetically modified tau-deficient animal models do not have A $\beta$ -induced spontaneous epileptiform activity, and show reversed excitation: inhibition imbalance and reduced vulnerability to chemically-induced seizures [38].

#### *Huperzine A as a therapeutic agent in AD*

Huperzine A (HupA) is an AChE inhibitor extracted from *Huperzia serrata*, a firmoss. HupA is a more potent and specific central nervous system (CNS) AChE inhibitor than other inhibitors such as donepezil, tacrine, and pyridostigmine in preclinical studies [39]. HupA has been used clinically to improve cognition and memory in AD and other forms of dementia in China. It is currently marketed as a memory enhancing dietary supplement in the US. In clinical trials, HupA has demonstrable cognition-improving capacity in AD patients and is not associated with any serious side effects in either this vulnerable patient population or in healthy volunteers [40,41]. HupA reduces A $\beta$  production and protects cells from A $\beta$ 1–42- or A $\beta$ 25–35-induced damage [42–45]. While HupA is also a weak N-methyl-D-aspartate glutamate receptor (NMDAR)

antagonist [46], its NMDAR inhibitory activity is not likely achieved at tolerable doses. In addition, as recently shown by our laboratory, HupA has potent antiepileptic effects in the rat pentylentetrazole (PTZ) model [47]. In light of these findings, HupA may have multiple benefits for mitigating the progression and AD symptoms.

#### *AChRs and HupA effects*

The 2 types of acetylcholinesterase receptors (AChRs) are nicotinic and muscarinic. They mediate fast and slow synaptic transmission, respectively. Muscarinic ACh receptors (mAChRs) are G-protein coupled and predominantly present in the parasympathetic nervous system. Nicotinic AChRs (nAChRs), neuronal- and muscle-type, are ion channels and have 5 subunits. Besides their roles in autonomic ganglia, nAChRs have a range of roles in neurotransmitter release, excitatory transmission, neurodevelopment, and plasticity throughout the CNS (primarily in cortex and hippocampus) [48]. In AD, the number of both mAChRs and nAChRs are decreased, in addition to loss of cholinergic neurons and the choline acetyltransferase enzyme in the basal forebrain [49]. This reduced cholinergic tone is most proximally related to cognitive impairment in AD [50].

HupA preferentially inhibits tetrameric AChE in the brain. Tetrameric AChE comprises the majority of membrane-bound AChE at the synapse, and hence HupA, unlike other AChE inhibitors, may have preferential synaptic effects [39,51]. Synaptic tetrameric AChE is the predominant AChE form in most brain regions where it co-localizes with other synaptic receptors, e.g.  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors, NMDARs, nAChRs and mAChRs [52]. The  $\alpha$ 7 subunit-containing nAChR ( $\alpha$ 7nAChR) has central anti-inflammatory effects in animal stroke and traumatic brain injury (TBI) models which suggest anti-seizure via a brain anti-inflammatory mechanism [53–55]. Moreover, GABAergic interneurons express abundant nAChR and thus enable AChE to mediate pyramidal cell activity [56]. Notably, the brain regions with both abundant tetrameric AChE and  $\alpha$ 7nAChR, such as the hippocampus and frontal cortex are those that are most affected by AD [39,57,58]. Hence, it is plausible that central HupA anti-seizure effects take place after indirect activation of  $\alpha$ 7nAChRs.

HupA may also modulate AD symptoms via mAChRs and  $\alpha$ 4 $\beta$ 2-subunit containing-nAChRs ( $\alpha$ 4 $\beta$ 2nAChRs). An in vitro study in rat hippocampal synaptosomes shows that GABA release is facilitated via  $\alpha$ 4 $\beta$ 2nAChRs [59]. Another in vivo study demonstrates  $\alpha$ 4 $\beta$ 2nAChRs increase GABAergic transmission in mouse corticolimbic neurons [60]. Another in vitro study also shows that mAChRs contribute to GABAergic inhibitory signaling in rat hippocampal neurons [61,62]. Taken together, these early preclinical data raise prospects for HupA to act indirectly via mAChRs and nAChRs to augment inhibitory tone and reduce cortical excitability in AD.

#### **Hypothesis**

Given that seizures in AD patients are difficult to control at tolerable doses of available AEDs, we hypothesize that HupA may be an effective and well-tolerated agent to both suppress seizures and improve cognition in AD by enhancing cholinergic and GABAergic signaling and mitigating AD-related neurotoxicity.

#### *HupA as an antiepileptic drug in AD*

HupA has acute anticonvulsant properties in the PTZ rat model [47] where it prophylaxes against seizures, enhances gamma oscillations on EEG cortical neurons from mice with mutant hAPP

which are associated with enhanced GABAergic cortical interneuron activity, and increases GABA-mediated cortical inhibition as measured by paired-pulse transcranial magnetic stimulation (ppTMS). Furthermore, inflammatory cytokines in brain probably mediate epileptogenic processes, and intervening with their actions are proposed to be potential targets for anticonvulsant drugs [55,63]. Several important mediators in this regard are tumor necrosis factor alpha (TNF- $\alpha$ ), interleukin 1-beta (IL-1 $\beta$ ), and nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B). In addition, along with A $\beta$ -mediated pathological changes, chronic neuroinflammation likely plays a role in AD pathogenesis [64]. Elevated levels of cytokines such as IL-1 $\beta$ , TNF- $\alpha$  and TGF- $\beta$  are found in postmortem AD brains, especially in late-stage disease [65]. Moreover, increase in IL-1 $\beta$ , TNF- $\alpha$  and microglial activation have been correlated in aged rat hippocampus which in turn potentially causes loss of GABAergic cells [66]. In line with these, HupA has been shown to decrease IL-1 $\beta$ , NF- $\kappa$ B and TNF- $\alpha$  levels in animal models [67–69]. Hence, HupA may exert sub-acute anticonvulsant and antiepileptic effects indirectly via cholinergic anti-inflammatory mechanisms regulated by nAChRs. Additionally, HupA may facilitate GABAergic signaling by NF- $\kappa$ B and IL-1 $\beta$  pathway suppression [63,70]. In parallel, preclinical studies show that A $\beta$  activates NF- $\kappa$ B pathway especially in hippocampus and prefrontal cortex, which subsequently causes induction of pro-inflammatory cytokines e.g. TNF- $\alpha$  and IL-1 $\beta$  [71]. By virtue of these plausible downstream effects, HupA may reverse the pro-inflammatory state in AD, which may also provide seizure protection.

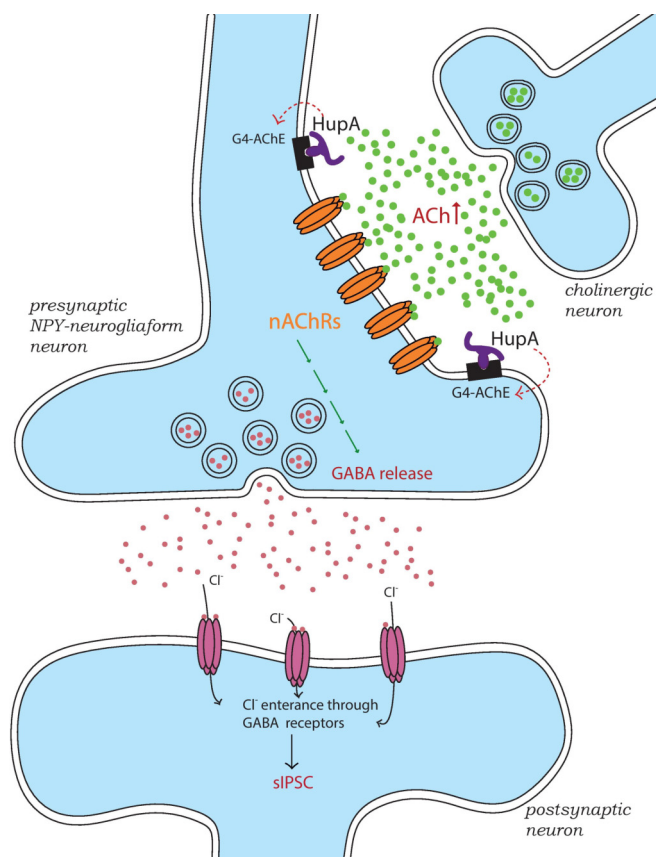
#### Can HupA prevent seizures in AD-associated hippocampal sclerosis?

Preclinical HS models show an upregulation of microglial cell markers and genes related to NF- $\kappa$ B pathways in epileptogenic tissue [72]. In addition, decrease in expression of receptor subtype genes such as that of GABA-A were argued to play roles in epileptogenesis in HS. Reduction in expression of neuropeptide Y (NPY) was also noted. This reduction contributes to hyperexcitability, given that intrahippocampal NPY administration suppresses seizures in rats and NPY-neurogliaform interneurons are inhibitory via slow, GABA-A receptor mediated inhibitory post synaptic currents in hippocampus and cortex (Fig. 1) [27,73,74].

HupA has central anti-inflammatory effects plausibly via suppressing NF- $\kappa$ B pathway and interleukin release from neurons and microglial cells. In addition, HupA facilitates GABAergic signaling whose mechanism is still unclear [57]. Since NPY-neurogliaform interneurons have dense cholinergic input, HupA may enhance GABAergic transmission through NPY pathways in the hippocampus where tetrameric AChE is abundant and HupA displays a preferential targeting effect [27,39,75]. Taken together, HupA may be hypothesized to provide anti-seizure activity in AD-associated HS via its cholinergic and GABAergic effects.

#### Can HupA reverse A $\beta$ - and hyperphosphorylated tau-related seizures?

Neuroigin-1 (NL1) is a transmembrane cell adhesion protein which helps organization and functioning of specifically excitatory synapses, ergo glutamatergic transmission and long-term potentiation [76]. NL1 has an extracellular domain homologous to AChE which interacts with A $\beta$  oligomers and stimulates A $\beta$  oligomerization [77,78]. An in vivo study suggests that NL1 overexpression causes an increased excitation:inhibition ratio in NL1-overexpressing transgenic mice, and another study reports that NL1 knockdown mice have less frequent and severe seizures [79,80]. We therefore hypothesize that interfering with NL1 related pathway by interacting with the AChE-like domain of NL1 with



**Fig. 1.** Inhibiting AChE, HupA may indirectly activate nAChRs on NPY-neurogliaform neurons. Thereby, increased GABAergic signaling may augment inhibitory tone and decrease seizure propensity. (NP: neuropeptide Y, G4-ACh: tetrameric acetylcholinesterase, nAChR: nicotinic acetylcholine receptor, sIPSC: slow inhibitory postsynaptic currents).

HupA may disrupt A $\beta$ -NL1 and prevent A $\beta$  oligomerization-induced excitotoxicity.

Reducing tau hyperphosphorylation and interfering with tau functioning suppresses seizures in numerous animal models [81–83]. In addition,  $\alpha$ 7nAChR activation alleviates tau-mediated neurodegeneration [84,85]. Moreover, donepezil, an AChE inhibitor, enhances protein phosphatase 2A (PP2A) activity, therefore reducing tau-induced neuronal viability and neurodegeneration [86]. Given that HupA is a potent CNS AChE inhibitor and has plausible effects via  $\alpha$ 7nAChRs, it may also prevent tau-related seizure predisposition via PP2A enhancement.

#### Evaluation of hypothesis

An effective and tolerable AED for patients with patients with AD remains a significant unmet medical need. Given the mechanisms of epileptogenesis in AD, HupA is a promising antiepileptic agent that may normalize cortical excitation:inhibition ratio, and may also have disease-modifying properties.

Clinical trials show that HupA is a safe and well-tolerable drug at doses up to 0.4 mg twice daily in patients with AD and vascular dementia without serious adverse effects, except for nausea and mild dizziness [87,88], which could potentially be mitigated by slow-release formulations. Thus, clinical trials where outcome measures include seizure and interictal spike counts as recorded on EEG, as well as clinical screens for drug side-effects, are realistic. Furthermore, methods like florbetaben positron emission tomography (PET) and cerebrospinal fluid A $\beta$ 1-42 or hyperphosphorylated

tau levels may be used to detect A $\beta$  burden, and track disease modification by HupA [89,90].

Another means of hypothesis testing may be in preclinical rodent experiments. Aged mice or rats, or transgenic animals expressing pathogenic APP gene variants are reasonable HupA study subjects [91,92]. In addition to measuring seizures by EEG, HupA-mediated modulation of cortical inhibitory tone can also be measured by ppTMS, a technique that is available in both humans and rodents which our group recently adapted to HupA research [47]. In this experiment, electromyography recordings from rat brachioradialis muscles were used to display motor evoked potentials (MEPs). It was found that the ratio of second-to-first MEP amplitudes were smaller after intraperitoneal HupA administration, suggesting increased cortical inhibition. That is, ppTMS can be used to detect CNS target engagement (GABA-A receptor modulation) by a range of pharmaceuticals, and we envision this technique to translate to human HupA trials. In parallel with in vivo animal studies, A $\beta$  levels may be assessed over a range of HupA doses and treatment schedules ex vivo by immunohistochemistry, ELISA, size exclusion chromatography, or other common laboratory methods [93–95].

## Conclusions

Unprovoked seizures affect approximately 20% of patients with AD, and a safe and well-tolerated antiepileptic treatment remains an unmet need in this population. While conventional AEDs may provide seizure control, their sustained use is associated with serious side effects such as lethargy, cognitive decline and susceptibility to falls. HupA, a potent cholinergic agent, has a favorable safety profile in the elderly, as supported by clinical trials in the AD population. In addition, HupA has acute anti-seizure effects in a rat preclinical model. Thus, HupA is a promising drug addressing both mnemonic and epileptic symptoms in patients with AD, without major, treatment-limiting, side-effects.

## Overview box

1. While AD is a risk factor for the development of epilepsy, current approved treatment options for AD only help to improve cognition and are neither effective AEDs nor disease-modifying. Moreover, available AEDs may worsen cognition and have other serious side effects including sedation and falls. HupA is an AChE inhibitor that has been shown to be safe and that possesses newly discovered antiepileptic, cognitive-enhancing, and disease-modifying properties.
2. Given that there is no proven method to reduce seizures, improve cognition, and impact disease progression in AD, the proposed hypothesis may lead to the demonstration that HupA fills a substantial gap in AD treatment.
3. Initial animal studies should aim to confirm that HupA suppresses seizures and enhances cognition in relevant AD models. Further studies could then investigate the related effects on A $\beta$  aggregation, preferably with transgenic animal models such as mutant chimeric human APP.

## References

- [1] McGleeson BM, Dynan KB, Passmore AP. Acetylcholinesterase inhibitors in Alzheimer's disease. *Br J Clin Pharmacol* 1999;48(4):471–80.
- [2] Parsons CG, Danysz W, Quack G. Memantine is a clinically well tolerated N-methyl-D-aspartate (NMDA) receptor antagonist—a review of preclinical data. *Neuropharmacology* 1999;38(6):735–67.
- [3] Matsunaga S, Kishi T, Iwata N. Memantine monotherapy for Alzheimer's disease: a systematic review and meta-analysis. *PLoS ONE* 2015;10(4):e0123289.

- [4] McShane R, Areosa Sastre A, Minakaran N. Memantine for dementia. *Cochrane Database Syst Rev* 2006(2):CD003154.
- [5] Mendez M, Lim G. Seizures in elderly patients with dementia: epidemiology and management. *Drugs Aging* 2003;20(11):791–803.
- [6] Hauser WA, Morris ML, Heston LL, Anderson VE. Seizures and myoclonus in patients with Alzheimer's disease. *Neurology*. 1986;36(9):1226–30.
- [7] Mendez MF, Catanzaro P, Doss RC, ARguello R, Frey 2nd WH. Seizures in Alzheimer's disease: clinicopathologic study. *J Geriatr Psychiatry Neurol* 1994;7(4):230–3.
- [8] Rao SC, Dove G, Cascino GD, Petersen RC. Recurrent seizures in patients with dementia: frequency, seizure types, and treatment outcome. *Epilepsy Behav* 2009;14(1):118–20.
- [9] Amatniek JC, Hauser WA, DelCastillo-Castaneda C, Jacobs DM, Marder K, Bell K, et al. Incidence and predictors of seizures in patients with Alzheimer's disease. *Epilepsia* 2006;47(5):867–72.
- [10] Palop JJ, Chin J, Mucke L. A network dysfunction perspective on neurodegenerative diseases. *Nature* 2006;443(7113):768–73.
- [11] Palop JJ, Chin J, Roberson ED, Wang J, Thwin MT, Bien-Ly N, et al. Aberrant excitatory neuronal activity and compensatory remodeling of inhibitory hippocampal circuits in mouse models of Alzheimer's disease. *Neuron* 2007;55(5):697–711.
- [12] Sanchez PE, Zhu L, Verret L, Vossel KA, Orr AG, Cirrito JR, et al. Levetiracetam suppresses neuronal network dysfunction and reverses synaptic and cognitive deficits in an Alzheimer's disease model. *Proc Natl Acad Sci USA* 2012;109(42):E2895–903.
- [13] Zaman SH, Parent A, Laskey A, Lee MK, Borchelt DR, Sisodia SS, et al. Enhanced synaptic potentiation in transgenic mice expressing presenilin 1 familial Alzheimer's disease mutation is normalized with a benzodiazepine. *Neurobiol Dis* 2000;7(1):54–63.
- [14] Zhang XZ, Li XJ, Zhang HY. Valproic acid as a promising agent to combat Alzheimer's disease. *Brain Res Bull* 2010;81(1):3–6.
- [15] Yao ZG, Liang L, Liu Y, Zhang L, Zhu H, Huang L, et al. Valproate improves memory deficits in an Alzheimer's disease mouse model: investigation of possible mechanisms of action. *Cell Mol Neurobiol* 2014;34(6):805–12.
- [16] Cumbo E, Ligor LD. Levetiracetam, lamotrigine, and phenobarbital in patients with epileptic seizures and Alzheimer's disease. *Epilepsy Behav* 2010;17(4):461–6.
- [17] Mbizvo GK, Dixon P, Hutton JL, Marson AG. The adverse effects profile of levetiracetam in epilepsy: a more detailed look. *Int J Neurosci* 2014;124(9):627–34.
- [18] Fleisher AS, Truran D, Mai JT, Langbaum JB, Aisen PS, Cummings JL, et al. Chronic divalproex sodium use and brain atrophy in Alzheimer disease. *Neurology* 2011;77(13):1263–71.
- [19] Bergey GK. Initial treatment of epilepsy: special issues in treating the elderly. *Neurology* 2004;63(10 Suppl 4):S40–8.
- [20] Hartikainen S, Lonnroos E, Louhivuori K. Medication as a risk factor for falls: critical systematic review. *J Gerontol A Biol Sci Med Sci* 2007;62(10):1172–81.
- [21] Paterniti S, Dufouil C, Alperovitch A. Long-term benzodiazepine use and cognitive decline in the elderly: the Epidemiology of Vascular Aging Study. *J Clin Psychopharmacol* 2002;22(3):285–93.
- [22] Gray SL, Dublin S, Yu O, Walker R, Anderson M, Hubbard RA, et al. Benzodiazepine use and risk of incident dementia or cognitive decline: prospective population based study. *BMJ* 2016;352:190.
- [23] Carreno M, Gil-Nagel A, Sanchez JC, Elices E, Serratos JM, Salas-Puig J, et al. Strategies to detect adverse effects of antiepileptic drugs in clinical practice. *Epilepsy Behav* 2008;13(1):178–83.
- [24] Perucca P, Gilliam FG. Adverse effects of antiepileptic drugs. *Lancet Neurol* 2012;11(9):792–802.
- [25] Palop JJ, Mucke L. Epilepsy and cognitive impairments in Alzheimer disease. *Arch Neurol* 2009;66(4):435–40.
- [26] Baraban SC. Neuropeptide Y and epilepsy: recent progress, prospects and controversies. *Neuropeptides* 2004;38(4):261–5.
- [27] Ibanez-Sandoval O, Tecuapetla F, Unal B, Shah F, Koos T, Tepper JM. A novel functionally distinct subtype of striatal neuropeptide Y interneuron. *J Neurosci* 2011;31(46):16757–69.
- [28] Lurton D, Cavalheiro EA. Neuropeptide-Y immunoreactivity in the pilocarpine model of temporal lobe epilepsy. *Exp Brain Res* 1997;116(1):186–90.
- [29] Reitz C. Alzheimer's disease and the amyloid cascade hypothesis: a critical review. *Int J Alzheimers Dis* 2012;2012:369808.
- [30] Bayer TA, Wirths O, Majtenyi K, Hartmann T, Multhaup G, Beyreuther K, et al. Key factors in Alzheimer's disease: beta-amyloid precursor protein processing, metabolism and intraneuronal transport. *Brain Pathol* 2001;11(1):1–11.
- [31] Qiu T, Liu Q, Chen YX, Zhao YF, Li YM. Abeta42 and Abeta40: similarities and differences. *J Pept Sci* 2015;21(7):522–9.
- [32] Minkeviene R, Rheims S, Dobszay MB, Zilberter M, Hartikainen J, Fulop L, et al. Amyloid beta-induced neuronal hyperexcitability triggers progressive epilepsy. *J Neurosci* 2009;29(11):3453–62.
- [33] Horvath A, Szucs A, Barcs G, Noebels JL, Kamondi A. Epileptic seizures in Alzheimer disease: a review. *Alzheimer Dis Assoc Disord* 2016.
- [34] Ittner LM, Ke YD, Delerue F, Bi M, Gladbach A, van Eersel J, et al. Dendritic function of tau mediates amyloid-beta toxicity in Alzheimer's disease mouse models. *Cell* 2010;142(3):387–97.
- [35] Iqbal K, Gong CX, Liu F. Hyperphosphorylation-induced tau oligomers. *Front Neurol* 2013;4:112.

- [36] Sen A, Thom M, Martinian L, Harding B, Cross JH, Nikolic M, et al. Pathological tau tangles localize to focal cortical dysplasia in older patients. *Epilepsia* 2007;48(8):1447–54.
- [37] Jellinger KA. Head injury and dementia. *Curr Opin Neurol* 2004;17(6):719–23.
- [38] Roberson ED, Halabisky B, Yoo JW, Yao J, Chin J, Yan F, et al. Amyloid-beta/Fyn-induced synaptic, network, and cognitive impairments depend on tau levels in multiple mouse models of Alzheimer's disease. *J Neurosci* 2011;31(2):700–11.
- [39] Zhao Q, Tang XC. Effects of huperzine A on acetylcholinesterase isoforms in vitro: comparison with tacrine, donepezil, rivastigmine and physostigmine. *Eur J Pharmacol* 2002;455(2–3):101–7.
- [40] Li YX, Zhang RQ, Li CR, Jiang XH. Pharmacokinetics of huperzine A following oral administration to human volunteers. *Eur J Drug Metab Pharmacokinet* 2007;32(4):183–7.
- [41] Xu SS, Cai ZY, Qu ZW, Yang RM, Cai YL, Wang GQ, et al. Huperzine-A in capsules and tablets for treating patients with Alzheimer disease. *Zhongguo Yao Li Xue Bao* 1999;20(6):486–90.
- [42] Lei Y, Yang L, Ye CY, Qin MY, Yang HY, Jiang HL, et al. Involvement of intracellular and mitochondrial abeta in the ameliorative effects of huperzine A against oligomeric Abeta42-Induced injury in primary rat neurons. *PLoS ONE* 2015;10(5):e0128366.
- [43] Gao X, Zheng CY, Yang L, Tang XC, Zhang HY. Huperzine A protects isolated rat brain mitochondria against beta-amyloid peptide. *Free Radic Biol Med* 2009;46(11):1454–62.
- [44] Peng Y, Jiang L, Lee DY, Schachter SC, Ma Z, Lemere CA. Effects of huperzine A on amyloid precursor protein processing and beta-amyloid generation in human embryonic kidney 293 APP Swedish mutant cells. *J Neurosci Res* 2006;84(4):903–11.
- [45] Zhang HY, Liang YQ, Tang XC, He XC, Bai DL. Stereoselectivities of enantiomers of huperzine A in protection against beta-amyloid(25–35)-induced injury in PC12 and NG108-15 cells and cholinesterase inhibition in mice. *Neurosci Lett* 2002;317(3):143–6.
- [46] Zhang YH, Chen XQ, Yang HH, Jin GY, Bai DL, Hu GY. Similar potency of the enantiomers of huperzine A in inhibition of [(3)H]dizocipine (MK-801) binding in rat cerebral cortex. *Neurosci Lett* 2000;295(3):116–8.
- [47] Gersner R, Ekstein D, Dhamne SC, Schachter SC, Rotenberg A. Huperzine A prophylaxis against pentylentetrazole-induced seizures in rats is associated with increased cortical inhibition. *Epilepsy Res* 2015;117:97–103.
- [48] Schaaf CP. Nicotinic acetylcholine receptors in human genetic disease. *Genet Med* 2014;16(9):649–56.
- [49] Lombardo S, Maskos U. Role of the nicotinic acetylcholine receptor in Alzheimer's disease pathology and treatment. *Neuropharmacology* 2015;96(Pt B):255–62.
- [50] Kihara T, Shimohama S. Alzheimer's disease and acetylcholine receptors. *Acta Neurobiol Exp (Wars)* 2004;64(1):99–105.
- [51] Xie HQ, Leung KW, Chen VP, Chan GK, Xu SL, Guo AJ, et al. PRiMA directs a restricted localization of tetrameric AChE at synapses. *Chem Biol Interact* 2010;187(1–3):78–83.
- [52] Skau KA, Shipley MT. Phenylmethylsulfonyl fluoride inhibitory effects on acetylcholinesterase of brain and muscle. *Neuropharmacology* 1999;38(5):691–8.
- [53] Han Z, Li L, Wang L, Degos V, Maze M, Su H. Alpha-7 nicotinic acetylcholine receptor agonist treatment reduces neuroinflammation, oxidative stress, and brain injury in mice with ischemic stroke and bone fracture. *J Neurochem* 2014;131(4):498–508.
- [54] Kelso ML, Oestreich JH. Traumatic brain injury: central and peripheral role of alpha7 nicotinic acetylcholine receptors. *Curr Drug Targets* 2012;13(5):631–6.
- [55] Vezzani A. Epilepsy and inflammation in the brain: overview and pathophysiology. *Epilepsy Curr* 2014;14(1 Suppl):3–7.
- [56] Buhler AV, Dunwiddie TV. Alpha7 nicotinic acetylcholine receptors on GABAergic interneurons evoke dendritic and somatic inhibition of hippocampal neurons. *J Neurophysiol* 2002;87(1):548–57.
- [57] Damar U, Gersner R, Johnstone JT, Schachter S, Rotenberg A. Huperzine A as a neuroprotective and antiepileptic drug: a review of preclinical research. *Expert Rev Neurother* 2016;1–10.
- [58] Maier DL, Hill G, Ding M, Tuke D, Einstein E, Gurley D, et al. Pre-clinical validation of a novel alpha-7 nicotinic receptor radiotracer, [(3)H] AZ11637326: target localization, biodistribution and ligand occupancy in the rat brain. *Neuropharmacology* 2011;61(1–2):161–71.
- [59] Zappettini S, Grilli M, Lagomarsino F, Cavallero A, Fedele E, Marchi M. Presynaptic nicotinic alpha7 and non-alpha7 receptors stimulate endogenous GABA release from rat hippocampal synaptosomes through two mechanisms of action. *PLoS ONE* 2011;6(2):e16911.
- [60] Maloku E, Kadriu B, Zhubi A, Dong E, Pibiri F, Satta R, et al. Selective alpha4beta2 nicotinic acetylcholine receptor agonists target epigenetic mechanisms in cortical GABAergic neurons. *Neuropsychopharmacology* 2011;36(7):1366–74.
- [61] Yi F, Ball J, Stoll KE, Satpute VC, Mitchell SM, Pauli JL, et al. Direct excitation of parvalbumin-positive interneurons by M1 muscarinic acetylcholine receptors: roles in cellular excitability, inhibitory transmission and cognition. *J Physiol* 2014;592(16):3463–94.
- [62] Gonzalez JC, Albinana E, Baldelli P, Garcia AG, Hernandez-Guijo JM. Presynaptic muscarinic receptor subtypes involved in the enhancement of spontaneous GABAergic postsynaptic currents in hippocampal neurons. *Eur J Neurosci* 2011;33(1):69–81.
- [63] Falip M, Salas-Puig X, Cara C. Causes of CNS inflammation and potential targets for anticonvulsants. *CNS Drugs* 2013;27(8):611–23.
- [64] Krstic D, Knuesel I. Deciphering the mechanism underlying late-onset Alzheimer disease. *Nat Rev Neurol* 2013;9(1):25–34.
- [65] Morimoto K, Horio J, Satoh H, Sue L, Beach T, Arita S, et al. Expression profiles of cytokines in the brains of Alzheimer's disease (AD) patients compared to the brains of non-demented patients with and without increasing AD pathology. *J Alzheimers Dis* 2011;25(1):59–76.
- [66] Gavilan MP, Revilla E, Pintado C, Castano A, Vizuete ML, Moreno-Gonzalez I, et al. Molecular and cellular characterization of the age-related neuroinflammatory processes occurring in normal rat hippocampus: potential relation with the loss of somatostatin GABAergic neurons. *J Neurochem* 2007;103(3):984–96.
- [67] Tian GX, Zhu XQ, Chen Y, Wu GC, Wang J. Huperzine A inhibits CCL2 production in experimental autoimmune encephalomyelitis mice and in cultured astrocyte. *Int J Immunopathol Pharmacol* 2013;26(3):757–64.
- [68] Wang ZF, Tang XC. Huperzine A protects C6 rat glioma cells against oxygen-glucose deprivation-induced injury. *FEBS Lett* 2007;581(4):596–602.
- [69] Sui X, Gao C. Huperzine A ameliorates damage induced by acute myocardial infarction in rats through antioxidant, anti-apoptotic and anti-inflammatory mechanisms. *Int J Mol Med* 2014;33(1):227–33.
- [70] Cogswell JP, Godlevski MM, Wisely GB, Clay WC, Leesnitzer LM, Ways JP, et al. NF-kappa B regulates IL-1 beta transcription through a consensus NF-kappa B binding site and a nonconsensus CRE-like site. *J Immunol* 1994;153(2):712–23.
- [71] Medeiros R, LaFerla FM. Astrocytes: conductors of the Alzheimer disease neuroinflammatory symphony. *Exp Neurol* 2013;239:133–8.
- [72] Vieira AS, de Matos AH, do Canto AM, Rocha CS, Carvalho BS, Pascoal VD, et al. RNA sequencing reveals region-specific molecular mechanisms associated with epileptogenesis in a model of classical hippocampal sclerosis. *Sci Rep* 2016;6:22416.
- [73] Mazarati A, Wasterlain CG. Anticonvulsant effects of four neuropeptides in the rat hippocampus during self-sustaining status epilepticus. *Neurosci Lett* 2002;331(2):123–7.
- [74] Faust TW, Assoum M, Shah F, Tepper JM, Koos T. Novel fast adapting interneurons mediate cholinergic-induced fast GABAA inhibitory postsynaptic currents in striatal spiny neurons. *Eur J Neurosci* 2015;42(2):1764–74.
- [75] English DF, Ibanez-Sandoval O, Stark E, Tecuapetla F, Buzsaki G, Deisseroth K, et al. GABAergic circuits mediate the reinforcement-related signals of striatal cholinergic interneurons. *Nat Neurosci* 2012;15(1):123–30.
- [76] Jedlicka P, Vnencak N, Krueger DD, Jungenitz T, Brose N, Schwarzacher SW. Neuroigin-1 regulates excitatory synaptic transmission, LTP and EPSP-spike coupling in the dentate gyrus in vivo. *Brain Struct Funct* 2015;220(1):47–58.
- [77] Dinamarca MC, Weinstein D, Monasterio O, Inestrosa NC. The synaptic protein neuroigin-1 interacts with the amyloid beta-peptide. Is there a role in Alzheimer's disease? *Biochemistry* 2011;50(38):8127–37.
- [78] Dinamarca MC, Di Luca M, Godoy JA, Inestrosa NC. The soluble extracellular fragment of neuroigin-1 targets Abeta oligomers to the postsynaptic region of excitatory synapses. *Biochem Biophys Res Commun* 2015;466(1):66–71.
- [79] Fang M, Wei JL, Tang B, Liu J, Chen L, Tang ZH, et al. Neuroigin-1 knockdown suppresses seizure activity by regulating neuronal hyperexcitability. *Mol Neurobiol* 2016;53(1):270–84.
- [80] Dahlhaus R, Hines RM, Eadie BD, Kannangara TS, Hines DJ, Brown CE, et al. Overexpression of the cell adhesion protein neuroigin-1 induces learning deficits and impairs synaptic plasticity by altering the ratio of excitation to inhibition in the hippocampus. *Hippocampus* 2010;20(2):305–22.
- [81] Li Z, Hall AM, Kelinske M, Roberson ED. Seizure resistance without parkinsonism in aged mice after tau reduction. *Neurobiol Aging* 2014;35(11):2617–24.
- [82] Gheyara AL, Ponnusamy R, Djukic B, Craft RJ, Ho K, Guo W, et al. Tau reduction prevents disease in a mouse model of Dravet syndrome. *Ann Neurol* 2014;76(3):443–56.
- [83] Jones NC, Nguyen T, Corcoran NM, Velakoulis D, Chen T, Grundy R, et al. Targeting hyperphosphorylated tau with sodium selenate suppresses seizures in rodent models. *Neurobiol Dis* 2012;45(3):897–901.
- [84] Del Barrio L, Martin-de-Saavedra MD, Romero A, Parada E, Egea J, Avila J, et al. Neurotoxicity induced by okadaic acid in the human neuroblastoma SH-SY5Y line can be differentially prevented by alpha7 and beta2\* nicotinic stimulation. *Toxicol Sci* 2011;123(1):193–205.
- [85] Bitner RS, Nikkel AL, Markosyan S, Otte S, Puttfarcken P, Gopalakrishnan M. Selective alpha7 nicotinic acetylcholine receptor activation regulates glycogen synthase kinase3beta and decreases tau phosphorylation in vivo. *Brain Res* 2009;1265:65–74.
- [86] Noh MY, Koh SH, Kim SM, Maurice T, Ku SK, Kim SH. Neuroprotective effects of donepezil against Abeta42-induced neuronal toxicity are mediated through not only enhancing PP2A activity but also regulating GSK-3beta and nAChRs activity. *J Neurochem* 2013;127(4):562–74.
- [87] Rafiq MS, Walsh S, Little JT, Behan K, Reynolds B, Ward C, et al. A phase II trial of huperzine A in mild to moderate Alzheimer disease. *Neurology* 2011;76(16):1389–94.
- [88] Xu ZQ, Liang XM, Juan W, Zhang YF, Zhu CX, Jiang XJ. Treatment with Huperzine A improves cognition in vascular dementia patients. *Cell Biochem Biophys* 2012;62(1):55–8.
- [89] Sabri O, Sabbagh MN, Seibyl J, Barthel H, Akatsu H, Ouchi Y, et al. Florbetaben PET imaging to detect amyloid beta plaques in Alzheimer's disease: phase 3 study. *Alzheimers Dement* 2015;11(8):964–74.

- [90] Funke SA, Birkmann E, Willbold D. Detection of Amyloid-beta aggregates in body fluids: a suitable method for early diagnosis of Alzheimer's disease? *Curr Alzheimer Res* 2009;6(3):285–9.
- [91] Hofrichter J, Krohn M, Schumacher T, Lange C, Feistel B, Walbroel B, et al. Sideritis spp. extracts enhance memory and learning in Alzheimer's beta-amyloidosis mouse models and aged C57Bl/6 mice. *J Alzheimers Dis* 2016.
- [92] Chan J, Jones NC, Bush AI, O'Brien TJ, Kwan P. A mouse model of Alzheimer's disease displays increased susceptibility to kindling and seizure-associated death. *Epilepsia* 2015;56(6):e73–7.
- [93] Ren W, Xu M, Liang SH, Xiang H, Tang L, Zhang M, et al. Discovery of a novel fluorescent probe for the sensitive detection of beta-amyloid deposits. *Biosens Bioelectron* 2016;75:136–41.
- [94] Randrianjatovo-Gbalou I, Marcato-Romain CE, Girbal-Neuhauser E. Quantification of amyloid fibrils using size exclusion chromatography coupled with online fluorescence and ultraviolet detection. *Anal Biochem* 2015;488:19–21.
- [95] Bruggink KA, Jongbloed W, Biemans EA, Veerhuis R, Claassen JA, Kuiperij HB, et al. Amyloid-beta oligomer detection by ELISA in cerebrospinal fluid and brain tissue. *Anal Biochem* 2013;433(2):112–20.